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Continuous purification of hepatitis C virus-like particles by multi-column chromatography

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Introduction

- VLPs present challenges for DSP, thus alternative strategies are desired;
- We evaluated semi-continuous, multicolumn chromatography techniques, that have the potential to improve process economics;
- A simple serial connection of two chromatographic columns directs the effluent of g one column to an adjacent one, capturing the breakthrough from the previous column and avoiding product loss;
- This simple setup modification overcomes the limitations of single-column processes by achieving saturation of the first column;
- It also benefits from the countercurrent flow between the mobile and the stationary phases, which optimizes the driving force for mass transfer throughout the overall trajectory of the two phases;

AIM: Evaluate continuous multicolumn chromatography for hepatitis C VLP purification

Materials & Methods

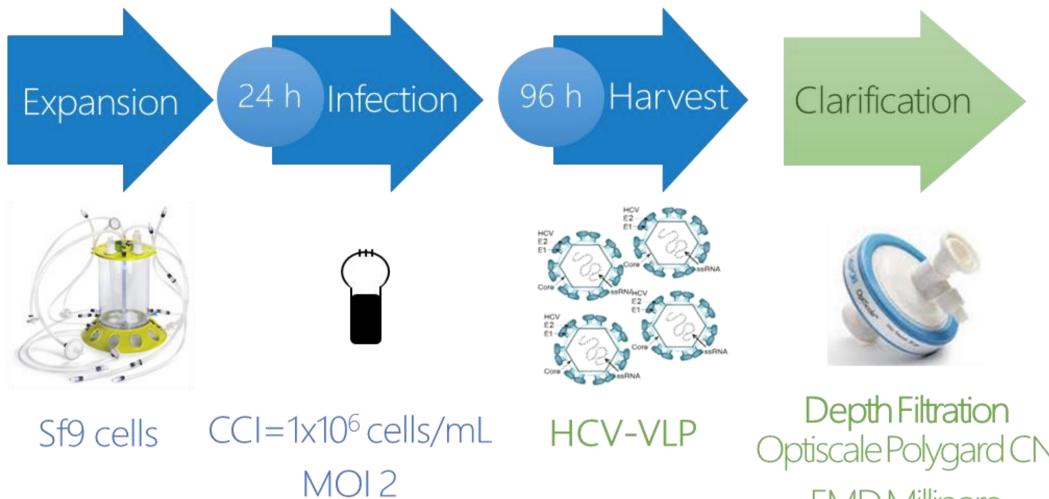


Figure 1: Steps and materials used for preparation of the HCV-VLP bulk

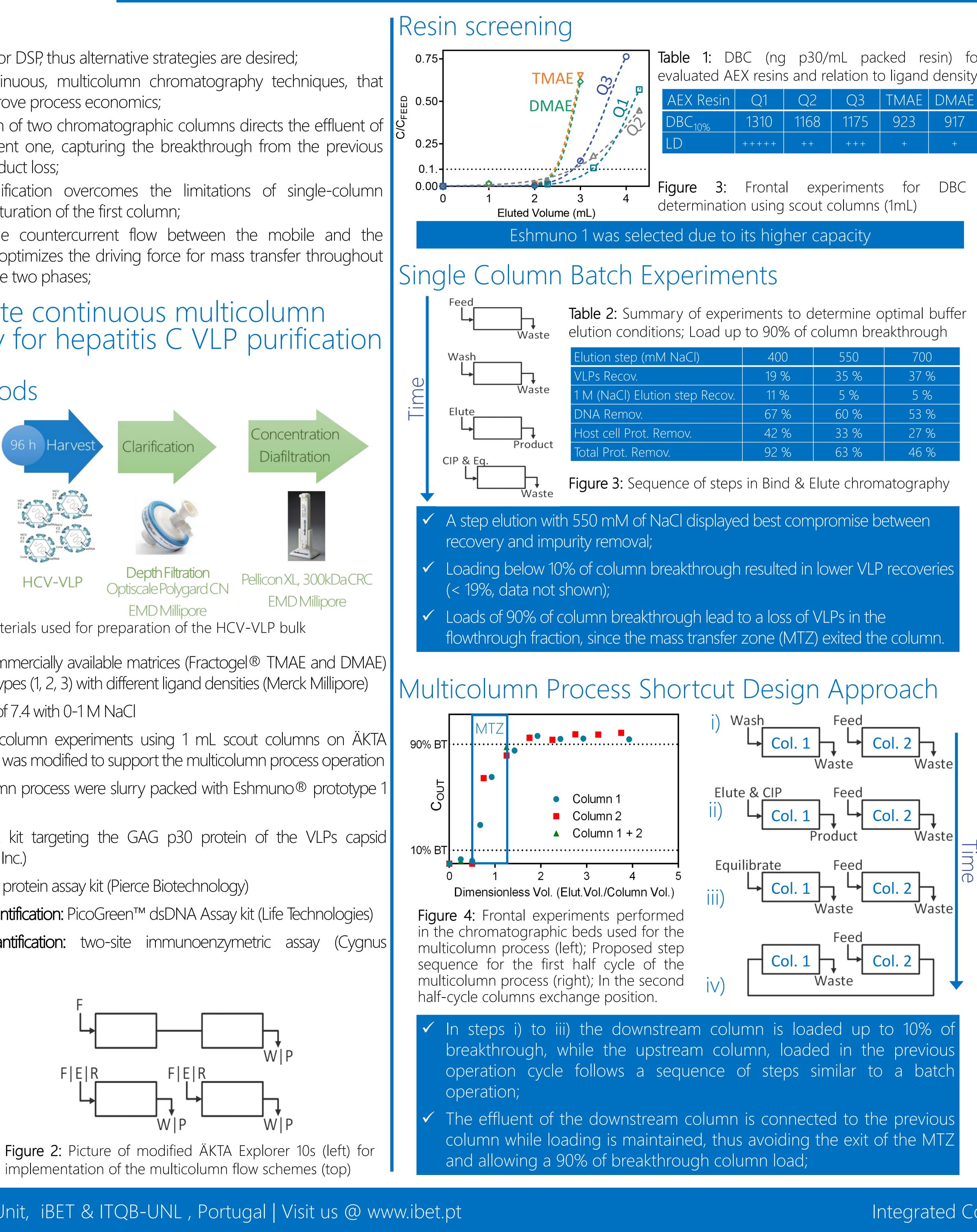
Anion exchange resins: two commercially available matrices (Fractogel® TMAE and DMAE) and three Eshmuno[®] Q prototypes (1, 2, 3) with different ligand densities (Merck Millipore) Buffers: 50 mM HEPES at a pH of 7.4 with 0-1 M NaCl

DBC determination for single-column experiments using 1 mL scout columns on ÄKTA Explorer 10s system. This system was modified to support the multicolumn process operation Columns used in the multicolumn process were slurry packed with Eshmuno® prototype 1 to a final bed volume of 4 mL

HCV-VLP quantification: ELISA kit targeting the GAG p30 protein of the VLPs capsid (QuickTiter™ MuLV, Cell Biolabs, Inc.)

Total Protein quantification: BCA protein assay kit (Pierce Biotechnology) Total double stranded DNA quantification: PicoGreen[™] dsDNA Assay kit (Life Technologies) host cell protein quantification: two-site immunoenzymetric assay (Cygnus Total Technologies, Inc.)





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Continuous purification of hepatitis C virus-like particles by multicolumn chromatography Ricardo J.S. Silva¹, Alex Xenopoulos², José P.B. Mota³, Cristina Peixoto^{1,4}, Manuel J.T. Carrondo^{1,5}, Paula M. Alves^{1,4}

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D	BC (r	ng	p30/r	mL pad	cked re	esin)	for
AEX resins and relation to ligand density							
•	$\bigcirc 1$		\frown	\bigcirc			

in	Q1	Q2	Q3	IMAE	DMAE
	1310	1168	1175	923	917
	+++++	++	+++	+	+

	400	550	700
	19 %	35 %	37 %
COV.	11 %	5 %	5 %
	67 %	60 %	53 %
	42 %	33 %	27 %
	92 %	63 %	46 %

Experimental Run

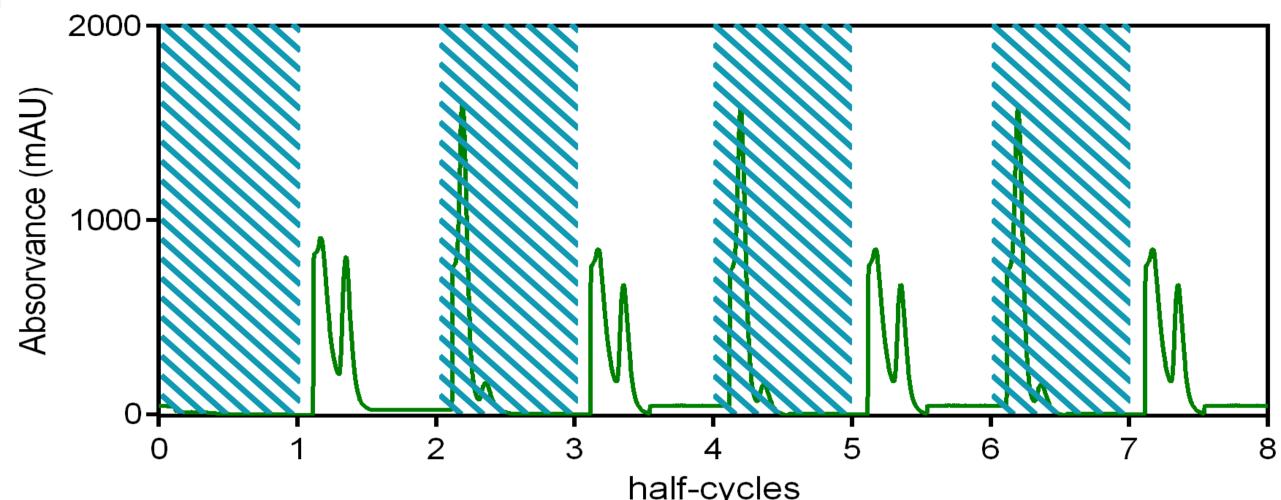
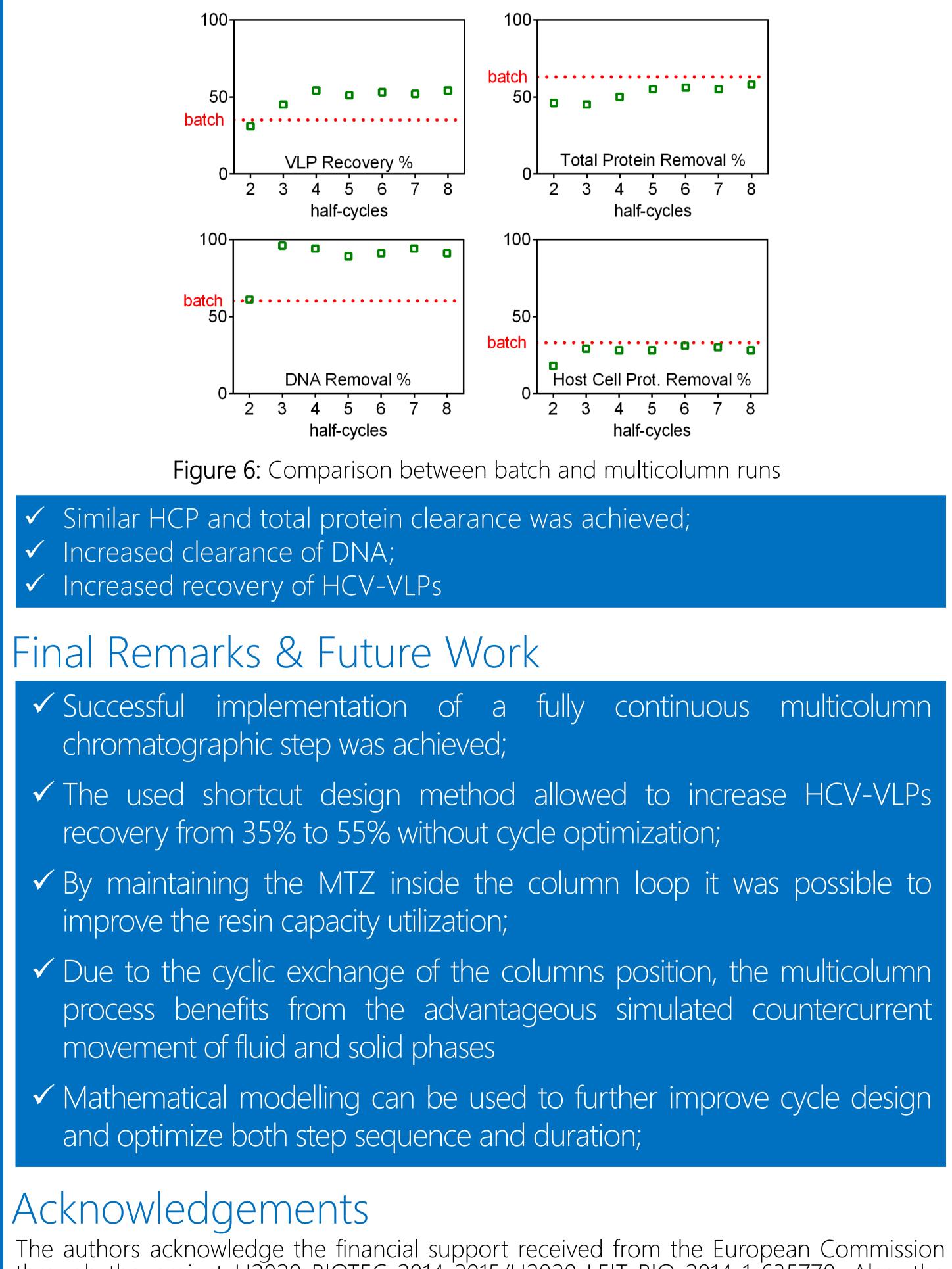


Figure 5: Experimental run of the proposed multicolumn process for continuous purification of HCV-VLPs; Shaded areas correspond to product collection in column 1; non-shaded areas correspond to the product collection in column 2;



material

COMPETE

4.2

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